

Development of early maturity maize hybrids for resistance to *Fusarium* and *Aspergillus* ear rots and their associated mycotoxins

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Abstract

Maize is mainly affected by two fungal pathogens, *Fusarium verticillioides* and *Aspergillus flavus*, causing *Fusarium* ear rot (FER) and *Aspergillus* ear rot (AER), respectively. Both fungi are of concern to stakeholders as they affect crop yield and quality, contaminating maize grains with the mycotoxins fumonisins and aflatoxins. The easiest strategy to prevent pre-harvest contamination by *F. verticillioides* and *A. flavus* is to develop maize hybrids resistant to FER and AER, as well as to their associated mycotoxins. The objective of this investigation was to test 46 F₁ hybrids, originated from different Italian, US and Canadian breeding groups, for these important traits and their agronomic performances. All hybrids were planted and artificially inoculated with toxigenic strains of *F. verticillioides* and *A. flavus* at two locations in 2017, and the best performing 17 out of 46 were also tested in 2018. Ear rots were present in all hybrids in 2017 and 2018, with percentages ranging from 6.50 to 49.50%, and 5.50 to 45.53%, for FER and AER, respectively. Seven hybrids (PC8, PC15, PC9, PC11, PC14, PC34 and PC17) presented the lowest levels of both diseases considering the overall locations and growing seasons, and three of these (PC8, PC11 and PC14) were also amongst the least mycotoxin contaminated hybrids in 2017. The inbred lines used in hybrid production may provide additional sources of resistance suitable in breeding programs targeting multiple pathogens and their mycotoxins.

Keywords: *Fusarium* ear rot, *Aspergillus* ear rot, resistance, mycotoxins, grain yield, maize hybrids

1. Introduction

Maize (*Zea mays* L.) is the most cultivated cereal grain in the world for yield and second for cultivated area (FAO, 2018). This cereal is a staple crop in Africa and in developing countries, while in developed ones is used mainly to feed livestock and for energy purposes. It is reported that in the near future the consumption of cereals will increase driven by an improved demand of cereals for animal feeding, which relies for 70% on maize (FAO, 2017).

Maize is affected by numerous fungal pathogens colonizing developing kernels and ears and causing ear rots. Many of these fungi are able to produce toxic secondary metabolites, known as mycotoxins that affect the quality and marketability of grains (Lanubile *et al.*, 2017a; Marin *et al.*, 2013; Morales *et al.*, 2018). Among these fungi,

two of the most relevant are *Fusarium verticillioides* and *Aspergillus flavus*. *F. verticillioides* (Sacc.) Nirenberg (synonym *Fusarium moniliforme* Sheldon, teleomorph *Gibberella moniliformis* Wineland) is the causal agent of stalk rot and *Fusarium* ear rot (FER) in maize, which can be found almost in all maize fields at harvest especially in humid and temperate climates nonetheless in tropical and subtropical countries (Lanubile *et al.*, 2017a; Morales *et al.*, 2018). *A. flavus*, the causal agent of *Aspergillus* ear rot (AER), is mainly a saprophyte and maize can be severely affected both in the field and during storage resulting in severe rotting of kernels and ears. The occurrence of AER is primarily in regions characterised by hot and dry climates (Lanubile *et al.*, 2014a) and is a main concern in Africa (Okoth *et al.*, 2017).

The presence of these two fungi in maize fields results in the contamination of the grain with mycotoxins. *F. verticillioides* is able to produce several toxins, such as moniliformins, fusarins and fumonisins, where the most relevant is fumonisin B₁, while *A. flavus* produces aflatoxins (Desjardins and Proctor, 2007; Marin *et al.*, 2013). From a toxicological perspective these mycotoxins are carcinogenic compounds responsible of acute and chronic disorders both in humans and animals (Lanubile *et al.*, 2014a, 2017a; Marin *et al.*, 2013; Okoth *et al.*, 2017; Szabo *et al.*, 2018; Warburton *et al.*, 2013).

An efficient solution to reduce ear rots and mycotoxin contamination is the development of host resistance, preferably in locally-adapted breeding materials. However, maize hybrids grown currently have insufficient resistance levels (Warburton *et al.*, 2013; Zila *et al.*, 2013). Many studies have focused on the research of resistance genetic markers for ear rot diseases (Gaikpa and Miedaner, 2019; Maschietto *et al.*, 2017; Robertson *et al.*, 2006; Warburton *et al.*, 2013; Zila *et al.*, 2013, 2014), in the selection of resistant inbred (Lanubile *et al.*, 2010; Reid *et al.*, 2009), in the development of adequate phenotyping techniques (Chungu *et al.*, 1996; Ju *et al.*, 2017; King and Scott, 1982; Lanubile *et al.*, 2014a; Septiani *et al.*, 2019; Stagnati *et al.*, 2019) and in the identification of low cost indicators of mycotoxin contamination (Morales *et al.*, 2018).

With the purpose to evaluate hybrid performances, the present work focused on the development of 46 hybrids crossing inbred lines developed in different breeding programs for broad resistance against ear rots. All hybrids were evaluated for resistance to FER and AER, mycotoxin accumulation and agronomic performances in two locations in 2017, and the best performing 17 out of 46 were also tested in 2018. The results showed that seven hybrids (PC8, PC15, PC9, PC11, PC14, PC34 and PC17) presented the lowest levels of both FER and AER diseases considering the overall locations and growing seasons. Such genetic material may be useful for maize breeders to manage ear rot diseases and mycotoxin contamination.

2. Materials and methods

Plant materials

Forty-six F₁ hybrids were produced and evaluated (Supplementary Table S1). Female lines used to produce F₁ seeds were selected for their resistance to *F. verticillioides* and belong to different breeding experiments: twenty-four and nineteen S₅ and S₃ lines, respectively, and three S₇ Recombinant Inbred Lines (RIL).

Selection of S₅ female lines

The breeding project to produce female inbred lines resistant to ear rots started by crossing 6 different Italian lines (named Lo) to 4 different US lines (Supplementary Table S1). Lines of US origin were selected as sources of resistance to mycotoxigenic fungi to improve the resistance level of Lo materials. Mp313E was developed in Mississippi from a direct self from Tuxpan and it was the first line released against *A. flavus* (Scott and Zummo, 1990); Mo18W was released in 1960s, from the cross of WF9 × Mo22, as resistant to *A. flavus* (Zuber, 1973); the inbred Mp420 was released as a source of resistance to kernel infection by *A. flavus* and it was developed by selfing within a cross between the inbred Mp1 and an S₃ line from the open pollinated variety Hill Yellow Dent; Mp307 was developed as natural restorer of Texas cytoplasmic male-sterility and as resistant genotype against *A. flavus* infection (<https://npgsweb.ars-grin.gov>). The F₁s between Lo and US lines were backcrossed (BC) to the female Lo parents. The resulting BC₁F₁s were selfed and the deriving S₁ plants were self-pollinated and evaluated for plant morphology and flowering time. The S₂ families were grown ear-to-row, evaluated for plant morphology and resistance to ear rots under natural conditions, and for each row, a single selfed-ear was harvested until the generation of S₅-derived inbred lines was obtained.

Selection of S₇ and S₃ female lines

S₇ female lines were derived from the cross of two lines having contrasting phenotypes for FER resistance: the resistant CO441, used as female, and the susceptible CO354, used as male. Both lines were achieved by the Eastern Cereal and Oilseed Research Centre, Agriculture and Agri-Food Canada (Ottawa, Canada) and maintained by sibling at the Department of Sustainable Crop Production in Piacenza (Italy). 188 S₇ lines were developed and phenotypically evaluated for *F. verticillioides* resistance (Maschietto *et al.*, 2017). From the cross of the resistant RIL 104 and 156, 19 S₃ lines were selected based on plant morphology and resistance to artificial infection, according to Maschietto *et al.* (2017).

Hybrid production

F₁s hybrids were produced in Hijuela, San Francisco de Mostazal, Chile. For each hybrid to be produced, four rows of female line and two of male were planted. Rows were 5 m long, spaced 80 cm apart each row and 1 m aisle. Field trials were managed according to the standard agricultural practices followed in the area. Hybrids were produced by hand pollination. Female parents were crossed to one out of 3 different male inbred lines CO430, CO433 and CO441, according to their cycle length. The CO430, CO433 and CO441 were developed for resistance to three different

fungal diseases *Gibberella* ear rot, FER and Common Smut, respectively (Reid *et al.*, 2009). The list of the hybrids produced is reported in Supplementary Table S1. Commercial hybrids belonging to FAO maturity 400 and 500 classes were used as checks during 2017 [P1114 (500) and SUM405 (400)] and 2018 [Kefieros (500), SUM405 (400) and P003 (400)] growing seasons.

Field experiments

Field trials were set up during two years, 2017 and 2018, in two locations: San Zenone al Lambro (SZ) (45°19'23.84"N – 9°22'8.09"E, Milan, Italy) and Tribiano (T) (45°24'31.63"N – 9°23'20.60"E; Milan, Italy). In 2017, fields were sown on 4 and 7 April, while in 2018, on 27 April and 11 June for SZ and T, respectively. Each plot consisted of 4 rows 5 m long, spaced 80 cm apart each row and 1 m aisle. Three random replicates were planted for each hybrid in each location. Field trials were managed according to standard agricultural practices followed in the area. Harvesting data were 11 and 12 September in 2017 and 21 September and 30 October in 2018 for SZ and T locations, respectively.

Hybrids were phenotypically evaluated according to the UPOV protocol CPVO TP/2/3. Agronomic traits such as plants per row, lodged and smashed plants were taken on the central rows of the plots. Plant and ear heights were taken on five plants per plot, and ear/plant ratios were calculated. At harvesting, grain yield (t/ha) was recorded and reported to a final density of 7.2 plants/m²; grain moisture (%) was determined as well. Silk date was also recorded on every plot as the number of days between sowing and silk emergence on 50% of the plants in a plot.

Inoculation, disease severity and mycotoxin evaluation

The primary ear of 5 plants per plot was inoculated with a spore suspension of *F. verticillioides* MPVP 294 (Maschietto *et al.*, 2017) and *A. flavus* MPVP A2092 (Lanubile *et al.*, 2017b; Mauro *et al.*, 2013), both originating from Italy. These strains are part of the culture collection of the Department of Sustainable Crop Production, Università Cattolica del Sacro Cuore of Piacenza, Italy. Inoculum for both strains was produced to a final concentration of 1×10⁶ spores/ml. Ears were inoculated according to the pin-bar inoculation method at 15 DAP (days after pollination) (Lanubile *et al.*, 2017a; Maschietto *et al.*, 2017). Inoculated ears were harvested at maturity, and 5 additional ears were taken as non-inoculated controls.

Disease severity was evaluated on hand-harvested ears that were air dried in greenhouse. FER severity was visually evaluated, assessing the percentage of the rotted surface of the ear, using a 7 point severity grid and assigning 1 for the absence of infection, and numbers from 2 to 7 according to the percentage level of the infection, where 2 = 1-3%, 3 =

4-10%, 4 = 11-25%, 5 = 26-50%, 6 = 51-75% and 7 = 76-100% (Maschietto *et al.*, 2017). AER was evaluated by using an 11 point scale to assess the percentage of the rotted surface of the ear, scoring 0 for the absence of the infection and numbers from 1 to 10 according to the percentage level of the infection, where 1 = 1-10%, 2 = 11-20%, 3 = 21-30%, 4 = 31-40%, 5 = 41-50%, 6 = 51-60%, 7 = 61-70%, 8 = 71-80%, 9 = 81-90% and 10 = 91-100%.

Mycotoxin analysis was carried out only on samples harvested and evaluated in 2017 growing season. Ears belonging to the same plot were bulked and shelled; a random sample of about 150-200 kernels was milled with a laboratory mill (Cyclotec™ 1093 Sample Mill, FOSS) using a 1 mm mesh. Special attention was taken in the milling procedure avoiding cross-contamination of kernels showing different disease levels.

Mycotoxin contamination was evaluated using the VICAM Fumo-V AQUA and Afla-V AQUA strips (Watertown, MA, USA), a fluorometric-immunocapture assay, following manufacturer's protocol. Values of fumonisins (B₁+B₂+B₃) and aflatoxins (B₁+B₂+G₁+G₂) given in the text are expressed in µg/kg.

Selection of F₁ hybrids for the second year field trial

After the first year of trials during 2017, F₁ hybrids were selected according to the mean values of ear rot severity. A 99% confidence interval was calculated using the Rmisc package (Hope, 2013) available in R software (R core team, 2017) based on the mean of the hybrids used in 2017 experiment. The limit to retain F₁s was 19% for FER and 17.4% for AER. F₁s having one or both values lower than the confidence limit were retained. Further selection was made discarding materials that were visually unsatisfactory. In Supplementary Table S1 the 17 F₁ hybrids indicated in bold were evaluated during both growing seasons.

Data analysis

Data manipulation and visualisation was performed with R software (R core team, 2017) and confidence intervals (CI) were calculated according to CI function (CI=0.95; Hope, 2013). The average FER and AER scores of all the five inoculated ears in each plot was calculated. Box-Cox transformations of phenotypic data (Morales *et al.*, 2018) were performed using the MASS package (Venables and Ripley, 2002) available in R. λ values for transformations were: 5.4 for days to silking, 0.2 for FER, 0.3 for AER, 0.5 for grain yield, -1.5 for grain moisture, 2 for plant height, 0.1 for ear height, -3.3 for lodged plant percentage and -0.5 for smashed plant percentage. For FER and AER under natural infection +1 constant was added to account for 0 before data transformation with $\lambda=-0.1$ (N-FER) and $\lambda=-8.2$ (N-AER). For lodged and smashed plant +1 constant

was added during percentage computation to correct for 0. Regarding fumonisins each sample was read three times and the mean of the three readings was computed. A 0.01 constant was added to account for 0. Values were Box-Cox transformed $\lambda = 0.311$. Aflatoxins were analysed only for hybrids that passed the ear rot threshold for the second year of cultivation in 2018. Each sample was read three times and a sample mean was subsequently calculated. A 0.01 constant was added to account for 0. Values were Box-Cox transformed with $\lambda = 0.3879$.

Correlation between traits were calculated. Two-ways ANOVA were conducted to test differences between hybrids and locations, with the exception of AER in San Zenone during 2018 (one-way ANOVA). Least Significance Difference (LSD) was calculated according to the *LSD.test* function available in the R package *agricolae* applying the Bonferroni correction (De Mendiburu, 2017).

3. Results and discussion

Ear rots under artificial infection

The present study examined the effects of artificial inoculation with *F. verticillioides* and *A. flavus* on different maize F_1 hybrids grown in San Zenone (SZ) and Tribiano (T) locations (North Italy) in 2017 and 2018, considering the severity of ear rot diseases, mycotoxin content and several agronomic traits. The full list of 46 F_1 hybrids employed in this work is reported in Supplementary Table S1. In 2018, a subset of 17 F_1 hybrids was further selected based on the highest grain yield values and lowest FER and AER incidence compared to the commercial hybrids used in 2017 experiment, and examined for the same traits reported before (Supplementary Table S1).

During the growing season 2017, the 46 maize F_1 hybrids evaluated at SZ and T differed significantly ($P \leq 0.001$) in their expression of FER symptoms (Table 1, Supplementary Table S2) and the mean value of severity was 22.47% with minimum and maximum values of 9.87% and 49.50%, respectively.

In 2018, no significant differences were reported among the 17 selected hybrids for both locations, with values of severity ranging from 6.50 to 22.17%, but locations and hybrid \times location interactions were significant ($P \leq 0.001$ and $P \leq 0.05$, respectively) (Table 2, Supplementary Tables S2 and S3). This finding could be explained considering that a lower number of hybrids was evaluated (17 out of 46) and only the best performing materials were chosen. Furthermore, in 2017 fields were sown almost at the same time reducing the effect of climatic differences (Supplementary Figure S1). Conversely, in 2018 the late sowing dates, especially for the field located in Tribiano, determined a shift of the plant flowering time and consequently of the artificial

inoculations, influencing the development of the fungus. Additionally, in 2018 more intense rainfalls were recorded from July to October during the maturity stage of maize (about 225 vs 155 mm in 2017) determining about 10% less of FER and AER infection (Table 2, Supplementary Figure S1).

Focusing on the different maize hybrid subgroups, the lowest FER percentage was observed for hybrids developed from the CO441 male parent during both growing seasons (18.4 and 12% for 2017 and 2018, respectively; Figure 1A). Indeed, six out of ten hybrids showing the highest resistance to FER at both locations and years derived from the same male parent (PC8, PC11, PC14, PC1, PC9 and PC10; Table 3).

Significant differences were revealed among hybrids, locations and hybrid \times location for AER in 2017 ($P \leq 0.05$), but not in 2018 (Tables 1 and 2, Supplementary Tables S2 and S3). The mean severity was 23.03% and 12.05% in 2017 and 2018, respectively. In 2018, AER at T location was not considered due to the sporadic presence of the disease symptoms and the resulting arduous assessing of phenotypes. This was a consequence of the delayed planting date and artificial infections performed at mid-end August, as already discussed for FER. During that period temperatures were not adequate for *A. flavus* development, which usually requires high temperature (37 °C) for optimal growth (O'Brian *et al.*, 2007).

According to the different hybrid subgroups, in 2017 hybrids developed from S_5 female lines and CO441 male parent showed weaker AER symptoms (Figure 1B), and among the most resistant hybrids seven were in common with trait resistance to FER (PC8, PC15, PC9, PC11, PC14, PC34 and PC17; Table 3). In contrast, less pronounced differences among female and male parents were observed in 2018.

CO441 is an inbred line with good resistance levels to FER (Lanubile *et al.*, 2010, 2013, 2014b; Maschietto *et al.*, 2017; Reid *et al.*, 2009). The desirable performances of this line were confirmed by our findings not only towards FER, but emerged also to AER, making CO441 as the most promising line to transfer resistance. A previous study on the resistance of hybrids and their parental lines described that the correlation between the performance of the hybrid was higher with the father line than with the mother line (Mesterhazy *et al.*, 2012). This finding is in line with our results confirming the prevalence of the male line in the resistance to ear rots.

CO430 and CO433 were reported as sources of resistance to FER as well (Maschietto *et al.*, 2016; Reid *et al.*, 2009). During hybrid evaluation it was observed that F_1 s from CO433 exhibited the unfavourable trait of protruding ears or exposed ear tips. This makes ears and kernels

Table 1. Mean, minimum and maximum values, and two-way analysis of variance (mean squares, MS), for 46 maize experimental hybrids and commercial FAO hybrids evaluated in San Zenone and Tribiano locations during 2017 growing season.^{1,2}

Traits	Experimental hybrids			Control hybrids			
	Mean	Minimum	Maximum	Mean	Hybrid (MS)	Location (MS)	Hybrid × Location (MS)
FER (%)	22.47	9.87	49.50	15.74	345.50 ^{***}	274.90 ^{ns}	186.10 ^{ns}
AER (%)	23.03	13.23	45.53	30.30	2.64 [*]	10.93 [*]	2.69 [*]
N-FER (%)	7.66	1.63	22.70	2.51	0.90 ^{***}	0.08 ^{ns}	0.63 ^{***}
N-AER (%)	0.28	0.00	2.50	0.27	0.002 [*]	0.0001403 ^{ns}	0.0017520 ^{ns}
Fumonisin content (mg/kg)	121.00	33.02	262.90	126.83	28.34 ^{ns}	162.91 ^{**}	26.78 ^{ns}
Aflatoxin content (µg/kg)	181.64	40.81	324.56	132.90	131.02 ^{***}	9.00 ^{ns}	38.74 ^{ns}
Grain yield (t/ha)	10.22	6.11	18.44	14.99	3.24 ^{***}	90.69 ^{***}	0.37 ^{***}
Grain moisture (%)	15.59	13.98	16.08	16.59	6.680e-06 ^{***}	2.088e-04 ^{***}	1.450e-06 ^{ns}
Silking (DAS)	77.7	73.5	82.83	79.40	1.992e+18 ^{***}	5.948e+16 ^{ns}	1.491e+17 ^{***}
Plant height (cm)	231.7	181.7	293.0	255.70	2.31 ^{***}	13.66 ^{***}	0.14 [*]
Ear height (cm)	107.02	79.83	136.5	115.42	0.12 ^{***}	2.20 ^{***}	0.0095 [*]
Stalk lodging (%)	0.29	0.00	2.28	0.20	0.000312 ^{ns}	0.02 ^{***}	0.000319 ^{ns}
Smashed (%)	6.55	0.29	23.15	2.55	0.28 ^{***}	0.006877 ^{ns}	0.18 [*]

¹ The following traits were considered: *Fusarium* ear rot (FER, %), *Aspergillus* ear rot (AER, %), and natural *Fusarium* and *Aspergillus* infection (N-FER and N-AER, %), fumonisins (B₁+B₂+B₃) and aflatoxins (B₁+B₂+G₁+G₂) content (µg/kg), grain yield (t/ha), grain moisture (%), days to silking (DAS), plant height (cm), ear height (cm), stalk lodging (%) and smashed plants (%).

² ns = not significant; *** $P \leq 0.001$; ** $P \leq 0.01$; * $P \leq 0.05$.

Table 2. Mean, minimum and maximum values, and two-way analysis of variance (mean squares, MS), for 17 maize experimental hybrids and commercial FAO hybrids evaluated in San Zenone and Tribiano locations during 2018 growing season.^{1,2}

Traits	Experimental hybrids			Control hybrids			
	Mean	Minimum	Maximum	Mean	Hybrid (MS)	Location (MS)	Hybrid × Location (MS)
FER (%)	12.88	6.50	22.17	11.25	109 ^{ns}	5674 ^{***}	142 [*]
AER (%)	12.05	5.50	25.00	11.20	2.67 ^{ns,3}	/	/
Grain yield (t/ha)	8.21	6.09	10.76	10.54	1.16 ^{***}	51.68 ^{***}	0.46 ^{ns}
Grain moisture (%)	17.68	16.20	21.15	19.76	0.0000033 ^{***}	0.0013822 ^{***}	0.0000015 [*]
Silking (DAS)	60.68	58.83	62.50	61.75	2.542e+16 ^{**}	1.137e+19 ^{***}	2.140e+16 [*]
Plant height (cm)	238.80	199.20	262.50	240.83	0.81 ^{***}	3.57 ^{***}	0.17 ^{***}
Ear height (cm)	128.30	109.20	140.8	129.20	0.03 ^{***}	1.28 ^{***}	0.0129 ^{**}
Stalk lodging (%)	0.00	0.00	0.00	0	3.992e-05 ^{ns}	2.241e-04 [*]	4.384e-05 ^{ns}
Smashed (%)	2.09	0.00	5.49	0.60	0.09 ^{ns}	7.15 ^{***}	0.08 ^{ns}

¹ The following traits were considered: *Fusarium* ear rot (FER, %), *Aspergillus* ear rot (AER, %), grain yield (t/ha), grain moisture (%), days to silking (DAS), plant height (cm), ear height (cm), stalk lodging (%) and smashed plants (%).

² ns = not significant; *** $P \leq 0.001$; ** $P \leq 0.01$; * $P \leq 0.05$.

³ Only for San Zenone location.

more exposed to birds, insect and wind dispersed spores (Santiago *et al.*, 2015) and for this reason this aspect may have contributed to increased susceptibility.

A significant moderate correlation was obtained between FER and AER severity only at SZ location in 2017 ($r=0.58$, $P \leq 0.01$; Figure 2A). Furthermore, a significant weak relationship was observed between the two experimental fields for FER in the same year ($r=0.30$, $P \leq 0.05$; Figure 2B).

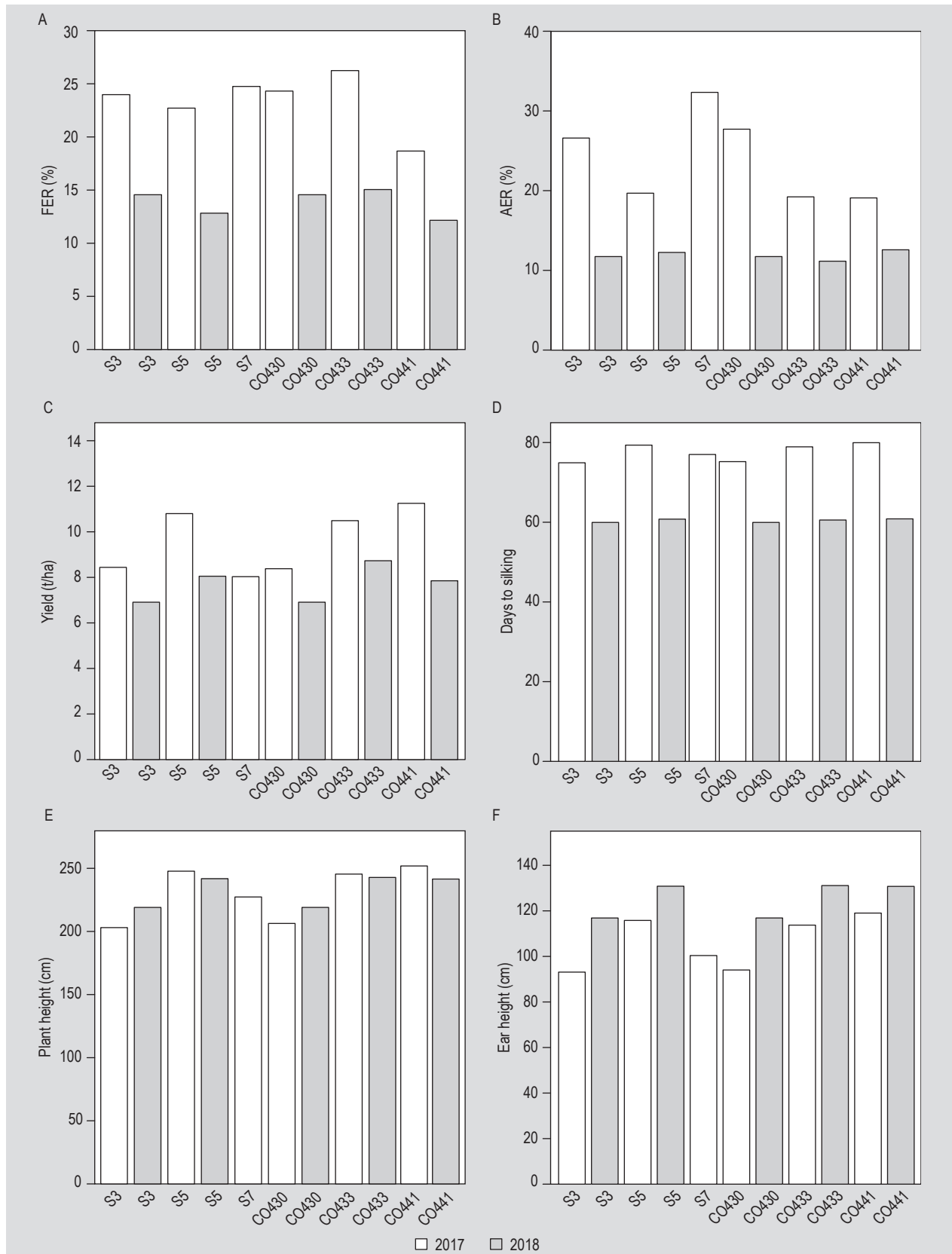


Figure 1. Mean values of *Fusarium* ear rot (FER, %) and *Aspergillus* ear rot (AER, %), yield (t/ha), days to silking, plant height (cm), and ear height (cm) over two locations according to maize hybrid subgroups during 2017 and 2018 growing seasons (white and grey histograms, respectively). The confidence intervals of the means for both years are: panel A 4.3, panel B 4.5, panel C 1.4, panel D 1.5, panel E 14.7, and panel F 8.6.

Table 3. Mean values for a subset of 10 maize experimental hybrids showing the lowest percentage (%) of *Fusarium* ear rot (FER) and *Aspergillus* ear rot (AER) in San Zenone (SZ) and Tribiano (T) locations during 2017 and 2018 growing seasons.¹

Trait	Hybrid	SZ_17	T_17	SZ_18	T_18	Overall mean
FER (%)	PC32	22.77 ^{bc}	8.87 ^{bc}	7.00 ^a	6.00 ^b	11.16
	PC8	7.92 ^{bc}	17.95 ^{bc}	6.00 ^a	13.00 ^b	11.22
	PC11	14.20 ^c	5.53 ^c	7.00 ^a	22.67 ^b	12.35
	PC17	12.15 ^{bc}	10.80 ^{bc}	4.67 ^a	26.33 ^b	13.49
	PC14	11.20 ^{bc}	17.93 ^{bc}	8.33 ^a	16.67 ^b	13.53
	PC1	13.20 ^{bc}	20.67 ^{bc}	7.00 ^a	14.33 ^b	13.80
	PC15	15.50 ^{abc}	21.07 ^{abc}	4.67 ^a	15.33 ^b	14.14
	PC9	10.87 ^{abc}	26.40 ^{abc}	6.33 ^a	13.33 ^b	14.23
	PC10	14.40 ^{bc}	17.67 ^{bc}	5.00 ^a	20.00 ^b	14.27
	PC34	19.07 ^{abc}	16.50 ^{abc}	7.67 ^a	15.33 ^b	14.64
AER (%)	PC16	8.60 ^{ab}	17.87 ^a	10.00	NA	12.16
	PC13	9.20 ^{ab}	23.89 ^a	7.50	NA	13.53
	PC8	8.67 ^{ab}	20.33 ^a	12.00	NA	13.67
	PC15	19.50 ^{ab}	12.62 ^a	9.33	NA	13.82
	PC9	10.33 ^{ab}	24.70 ^a	8.33	NA	14.46
	PC11	20.07 ^{ab}	13.87 ^a	10.00	NA	14.64
	PC12	16.20 ^{ab}	19.07 ^a	9.00	NA	14.76
	PC14	21.50 ^{ab}	19.25 ^a	7.00	NA	15.92
	PC34	29.33 ^{ab}	13.40 ^a	8.67	NA	17.13
	PC17	19.70 ^{ab}	19.67 ^a	14.00	NA	17.79

¹ Means followed by the same letter in the column do not differ significantly at the 0.05 probability level based on Bonferroni test.

The absence of correlation between the two diseases and the two locations in 2018 has to be most likely imputed to the late and different sowing times that resulted in postponed fungal infections, thus penalising their growth.

Identification of the traits that contribute to resistance to both FER and AER is an important task in breeding programs. Genotypes showing resistance to both *A. flavus* and *F. verticillioides* are known (Henry *et al.*, 2009) and the correlation between resistances against the two fungi was previously reported (Guo *et al.*, 2017; Robertson-Hoyt *et al.*, 2007; Rose *et al.*, 2017). Quantitative trait loci (QTL) associated to different ear rot pathogens suggest that these traits may be genetically linked (Gaikpa and Miedaner, 2019). In a previous study analysing the defence responses towards different FER-causing *Fusarium* spp. and *A. flavus*, an overlap of genes and enzymatic pathways was observed following the infection with these pathogens (Lanubile *et al.*, 2015), supporting the feasibility of selecting materials against multiple fungi at the same time.

Ear rots under natural infection

In 2017, susceptibility to natural infection caused by *F. verticillioides* and *A. flavus* was measured and F₁ hybrids significantly differed in Tribiano and San Zenone for both N-FER ($P \leq 0.001$) and N-AER ($P \leq 0.05$) (Table 1; Supplementary Table S2). Hybrid \times location interaction was significant ($P \leq 0.001$) for N-FER too (Table 1; Supplementary Table S2). The range of N-FER was from 1.63 to 22.70% with a mean of 7.66%. Natural AER was detected only in 16 hybrids with a maximum value of 2.67% and an average of 0.28%. Interestingly, strong and moderate correlations were observed in T ($r=0.73$, $P \leq 0.01$) and SZ ($r=0.55$, $P \leq 0.01$) between artificial and natural susceptibility of hybrids to FER. A weaker ($r=0.28$), but significant ($P \leq 0.05$) correlation was determined between the two locations for N-FER. No significant correlations were found for N-AER between T and SZ experimental fields. Natural infection of FER and AER was not measured in 2018 due to the almost complete absence of both fungi.

Up to sixteen species of *Fusarium* can coexist in field (Dorn *et al.*, 2011), and for the same species multiple strains can be present and not adequately controllable (Mesterhazy *et al.*, 2012). Additionally, natural infections are not always adequate and uniform, and for these reasons they should not be considered for selection program. Interestingly, significant correspondence between the results of resistance to FER after natural and artificial inoculations were observed in this study. Despite genotype differences are much wider under artificial inoculation, many breeders use natural infection pressure for the selection of disease resistant genotypes (Balconi *et al.*, 2014; Mesterhazy *et al.*, 2012). Our results evidenced the validity of the two tests both advisable for resistance breeding purposes.

Mycotoxin contamination

In 2017, artificially inoculated F₁ maize hybrids were evaluated for total fumonisin (B₁+B₂+B₃) and aflatoxin (B₁+B₂+G₁+G₂) content (Table 1, Supplementary Table S2). All samples were contaminated with both mycotoxins, exceeding the European legislation-recommended level in unprocessed maize of 4 mg/kg for fumonisins and 0.005 µg/kg for aflatoxins. Fumonisin levels varied among hybrids with total fumonisin concentrations averaging 121 mg/kg (ranging from 33.02 to 262.90 mg/kg) and total aflatoxins averaging 181.64 µg/kg (ranging from 40.81 to 324.56 µg/kg). Overall, differences in fumonisin contamination were not significant among hybrids, but they were for locations ($P \leq 0.001$; Table 1). Conversely, hybrids varied significantly for the aflatoxin content ($P \leq 0.001$; Table 1).

The less contaminated hybrids were PC17, PC44 and PC34 for fumonisins, whereas PC9, PC8 and PC20 were less contaminated with aflatoxins (Table 4). Interestingly, PC8,

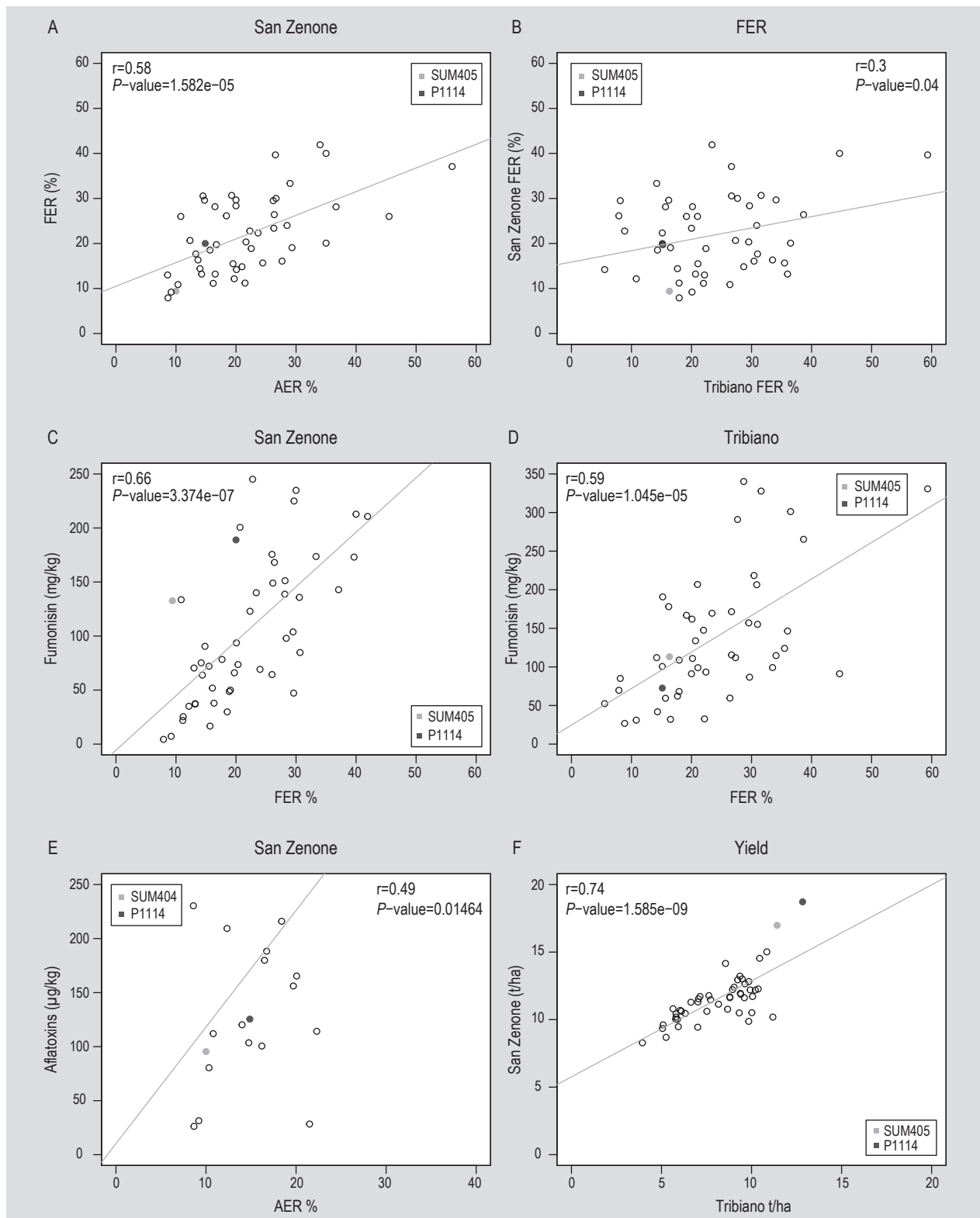


Figure 2. Correlations between traits during the 2017 growing season. (A) Percentage of *Fusarium* ear rot (FER) and *Aspergillus* ear rot (AER) at San Zenone location; (B) percentage of FER between San Zenone and Tribiano locations; percentage of FER and fumonisins (mg/kg) at San Zenone and Tribiano locations (C and D, respectively); (E) percentage of AER and aflatoxins ($\mu\text{g}/\text{kg}$) at San Zenone location; (F) grain yield (t/ha) between San Zenone and Tribiano locations; (G) percentage of FER and grain yield (t/ha) at San Zenone location; (H) percentage of AER and grain yield (t/ha) at San Zenone location; grain yield (t/ha) and days to silking (DAS) at San Zenone and Tribiano locations (I and L, respectively); (M) percentage of FER and DAS at San Zenone location; and (N) percentage of AER and DAS at San Zenone location.

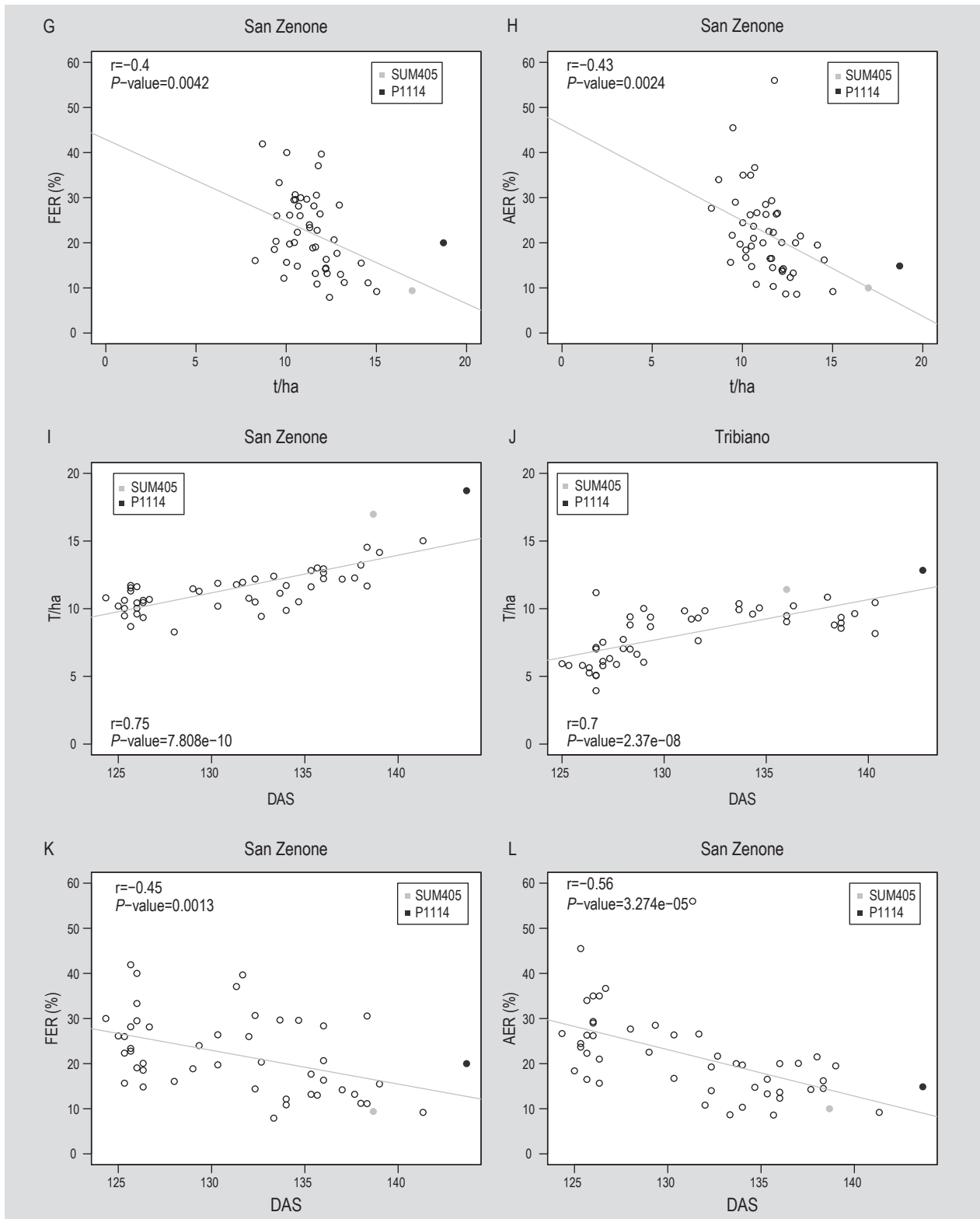


Figure 2. Continued.

PC10, PC11 and PC14 were among the most resistant hybrids to both mycotoxins and PC8, PC11 and PC14 showed also elevated resistance to FER and AER (Tables 3 and 4). Once again, the best performers were hybrids derived from the male parent CO441 and female lines S_5 coherently with previous findings regarding FER and AER. These three hybrids may be favourable sources of resistance in breeding efforts to reduce the presence of both diseases and their associated mycotoxins.

Good correlations between resistance to FER and accumulation of fumonisins was observed for both SZ ($r=0.66$, $P\leq 0.01$; Figure 2C) and T ($r=0.57$, $P\leq 0.01$; Figure 2D). Moreover, a moderate correlation of 0.49 was observed between AER severity and aflatoxin content in the location of SZ ($P\leq 0.01$; Figure 2E), suggesting that climatic conditions at Tribiano seemed more unfavourable for *A. flavus* growth and aflatoxin production, probably due to the more severe rainfalls and lower temperatures recorded in this location (Supplementary Figure S1).

Although these experiments indicated a positive overall relationship between ear rots and subsequent mycotoxin contamination (Figure 2), some hybrids did not follow this

Table 4. Mean values for a subset of 10 maize hybrids showing the lowest content of fumonisins ($B_1+B_2+B_3$) and aflatoxins ($B_1+B_2+G_1+G_2$) expressed in mg/kg and $\mu\text{g/kg}$, respectively, in San Zenone (SZ) and Tribiano (T) during 2017 growing season.¹

Trait	Hybrid	SZ_17	T_17	Overall mean
Fumonisin (mg/kg)	PC17	35.03 ^a	31.01 ^{ab}	33.02
	PC44	29.82 ^a	41.85 ^{ab}	35.84
	PC34	49.94 ^a	32.06 ^{ab}	41.00
	PC14	25.33 ^a	68.09 ^{ab}	46.71
	PC16	70.41 ^a	32.65 ^{ab}	51.53
	PC8	4.35 ^a	108.83 ^{ab}	56.59
	PC10	63.86 ^a	62.17 ^{ab}	63.02
	PC11	75.21 ^a	52.44 ^{ab}	63.82
	PC5	37.86 ^a	99.16 ^{ab}	68.51
	PC26	16.66 ^a	124.02 ^{ab}	70.34
Aflatoxins ($\mu\text{g/kg}$)	PC9	80.44 ^b	1.17 ^b	40.81
	PC8	26.22 ^{ab}	57.50 ^{ab}	41.86
	PC20	112.00 ^{ab}	44.67 ^{ab}	78.33
	PC14	28.33 ^{ab}	134.33 ^{ab}	81.33
	PC32	114.17 ^{ab}	91.45 ^{ab}	102.81
	PC13	31.44 ^{ab}	180.67 ^{ab}	106.06
	PC10	120.22 ^{ab}	100.00 ^{ab}	110.11
	PC11	165.33 ^{ab}	56.33 ^{ab}	110.83
	PC12	100.56 ^{ab}	156.56 ^{ab}	128.56
	PC25	103.56 ^{ab}	168.78 ^{ab}	136.17

¹ Means followed by the same letter in the column do not differ significantly at the 0.05 probability level based on Bonferroni test.

trend and results should be considered with caution because disease development and toxin accumulation were not uniform at all locations and years. Previous studies reported that differences on fumonisin accumulation existed for materials with the same disease severity (Mesterhazy *et al.*, 2012) and correlations between mycotoxins and fungal growth were variable (Munkvold *et al.*, 2003). Conversely, several works found good correlations (more than 0.85) between these two traits (Hung and Holland, 2012; Robertson *et al.*, 2006). Recently, Maschietto *et al.* (2017) reported a relatively high phenotypic correlation between FER and the presence of fumonisins. Moreover, they identified eight common QTL for these two traits further reinforcing the hypothesis of common resistance maize genetic mechanisms. Similar contrasting data were described for resistance to AER and aflatoxins. These two traits were reported to be highly correlated by Robertson-Hoyt *et al.* (2007). In another study, a strong genetic correlation between the AER and the aflatoxin concentration was recorded only in one location out of the four evaluated (Okoth *et al.*, 2017). Furthermore, Rose *et al.* (2017) identified two lines previously described as resistant to aflatoxin accumulation showing resistance to FER and fumonisin production.

Mycotoxin analysis was not carried out on samples harvested and evaluated in 2018 growing season due to the more fluctuating climatic conditions that caused a lower incidence of both ear rots. Additional in-depth analysis is required to elucidate the potential mechanism behind this phenomenon.

Agronomic performances of hybrids

The agronomic performances of the 46 F_1 hybrids are shown in Tables 1 and 2. Significant differences among hybrids were observed for all the agronomic traits considered in 2017 and 2018 for both locations ($P\leq 0.001$), with the exception of stalk lodging in both growing seasons and the percentage of smashed plants in 2018 (Tables 1 and 2, Supplementary Tables S2 and S3). Locations were significantly different for all traits in both years ($P\leq 0.001$), except DAS and percentage of smashed plants in 2017 (Tables 1 and 2, Supplementary Tables S2 and S3). Furthermore, hybrids \times location interaction was significant for almost all characters considered, excluding the percentage of stalk lodging in both years, grain moisture in 2017, and grain yield and percentage of smashed plants in 2018 (Tables 1 and 2, Supplementary Tables S2 and S3).

During hybrid development, grain yield represents a driving factor of breeding purposes. Average productions of 11.92 t/ha and 8.53 t/ha in 2017, and 10.05 t/ha and 6.38 t/ha in 2018 were obtained in SZ and T, respectively (Tables 1 and 2, Supplementary Table S2). Despite the highly significant good correlation observed for this trait

between the two locations in 2017 ($r=0.74$, $P\leq 0.01$; Figure 2F), undeniably, San Zenone and the year 2017 represented the most favourable environment and growing season for the evaluation of the F_1 hybrids and the expression of their yield performances. Indeed, moderate negative correlations were described only for SZ between susceptibility to both ear rots and grain yield in 2017 ($r=-0.40$, $P\leq 0.01$ and $r=-0.43$, $P\leq 0.01$ for FER and AER, respectively; Figure 2G and 2H), suggesting that the best performing hybrids exhibited the lower levels of FER and AER diseases. According to different hybrid groups, yield potentials were higher for hybrids deriving from CO441 and CO433 male parents and S_5 female lines (Figure 1C), which had longer DAS compared to S_3 and S_5 lines (Supplementary Table S2). As expected, a strong positive correlation was observed between DAS and grain yield for both San Zenone ($r=0.75$, $P\leq 0.01$) and Tribiano ($r=0.69$, $P\leq 0.01$) (Figure 2I and 2L).

DAS ranged from 73 to 83 days during 2017 and from 59 to 62 days during 2018 (Tables 1 and 2, Supplementary Table S2). Wider differences in DAS among hybrids were observed in 2017 (11 days) than 2018 (3 days), as a consequence of late planting for experimental fields. Delayed planting shortened the effective growing season, influencing flowering and fungal development as well (Nielsen *et al.*, 2002). Not particular differences were observed among the different hybrid groups for DAS, but in general it resulted higher in 2017, as previously described (Figure 1D). Moderately negative correlations were detected between DAS and FER ($r=-0.45$, $P\leq 0.01$) and DAS and AER ($r=-0.56$, $P\leq 0.01$) at SZ location in 2017 (Figure 2M and 2N), whereas no correlations were observed in Tribiano. It was previously described that late maturing inbreds and hybrids were potentially more at risk of severe ear rots (Battilani *et al.*, 2008; Eller *et al.*, 2008; Lanubile *et al.*, 2011; Santiago *et al.*, 2015). Instead, an opposite trend derived from our findings. Similarly, negative correlations were detected between silking dates and severity to *F. verticillioides* and *Fusarium graminearum* ear rots with later genotypes being more resistant (Mesterhazy *et al.*, 2012).

At harvesting, the percentage of grain moisture was not uniform among hybrids, ranging from 13.98 to 16.08% in 2017 and 16.20 and 21.15% in 2018 (Tables 1 and 2, Supplementary Tables S2 and S3). Grain is normally harvested at a moisture content of 20%, followed by artificial drying of the grain stock to avoid the presence of ear rots caused by *F. verticillioides*, *F. graminearum* and *A. flavus* and the resulting presence of mycotoxins. The rate of dry down of hybrids is a consequence of the genetic background of the material and is relevant for fungal infections and micotoxin accumulation. A previous study concerning dynamic of water activity and humidity of maize kernels revealed how 'slow dry down' hybrids were more prone to fumonisin accumulation, irrespective of their maturity class (Battilani *et al.*, 2011).

The mean of plant and ear height varied significantly in F_1 hybrids at both locations and years ($P\leq 0.001$) (Tables 1 and 2, Supplementary Tables S2 and S3). The average of both traits was higher in 2018 than in 2017, with mean values of 231.7 cm and 107.02 cm in 2017 and 238.8 cm and 128.3 cm in 2018 for plant and ear height, respectively. This finding could be due to the selection of hybrids carried out after the first year of trials in 2017, where the F_1 hybrids with higher values of ear rot severity and visually unsatisfactory were discarded. This was in line with the susceptibility to lodging and the percentage of smashed plants observed in the two years, reported to a greater extent in 2017, whereas the percentage of lodging was not recorded in 2018 at all (Tables 1 and 2, Supplementary Tables S2 and S3). As already come to light for the other traits, ear and plant height were higher for hybrids deriving from CO441 and CO433 male parents and S_5 female lines for both years, but more markedly for the year 2018 where the best performing 17 hybrids were chosen (Figure 1E and 1F). Previous studies reported that plant height, ear height, grain moisture and also, plant stand ability, had highly significant genetic and phenotypic direct effects on grain yield (Adu *et al.*, 2016; Filipovic *et al.*, 2014). Selection of hybrids having the best yield related traits represents an added value for the most disease and mycotoxin resistant materials identified in this study.

4. Conclusions

A major concern for global food and feed security is represented by the contamination of maize with *Fusarium* spp., *Aspergillus* spp. and their associated mycotoxins. Breeding for resistance using inbred lines and hybrids with high level of resistance to *F. verticillioides*/fumonisins and *A. flavus*/aflatoxins is still viewed to be one of the best approaches to mitigate this issue in maize. In this study, we identified seven hybrids (PC8, PC15, PC9, PC11, PC14, PC34 and PC17) resistant to both FER and AER diseases considering the overall locations and growing seasons, and three of these (PC8, PC11 and PC14) were also amongst the least contaminated hybrids in 2017. In the current study the identification of new hybrids and inbred lines with resistance to kernel infection by *F. verticillioides* and *A. flavus* provide additional sources of resistance that could be exploited in other breeding programs. This evidence is substantiated by the results on single crosses using parental lines that have been identified as resistant. Such genotypes can be used to develop early-medium maturity class of maize hybrids for use in areas of the country likely to have ear rot issues.

Supplementary material

Supplementary material can be found online at <https://doi.org/10.3920/WMJ2019.2554>.

Figure S1. Climatic variables represented by minimum, mean and maximum temperatures and rainfall verified at San Zenone and Tribiano locations during 2017 and 2018 years.

Table S1. List of hybrids, their origin, days to silking and to anthesis for female and male lines, respectively.

Table S2. Phenotypic data collected from the 46 F₁ hybrids and the four commercial hybrids evaluated during 2017 and 2018 years at San Zenone and Tribiano locations.

Table S3. Least significance difference analysis of the phenotypic traits from the 46 F₁ hybrids and the four commercial hybrids evaluated during 2017 and 2018 years at San Zenone and Tribiano locations.

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Conflict of interest

The authors declare no conflict of interest.

References

- Adu, G.B., Akromah, R., Abdulai, M.S., Obeng-Antwi, K., Alidu, H. and Tengan, K.M.L., 2016. Trait association for improved grain yield of extra-early maturing maize hybrids evaluated in the forest and transitional zones of Ghana. *Australian Journal of Crop Science* 10: 1127-1135.
- Balconi, C., Berardo, N., Locatelli, S., Lanzanova, C., Torri, A. and Redaelli, R., 2014. Evaluation of ear rot (*Fusarium verticillioides*) resistance and fumonisin accumulation in Italian maize inbred lines. *Phytopathologia Mediterranea* 53: 14-26.
- Battilani, P., Formenti, S., Ramponi, C. and Rossi, V., 2011. Dynamic of water activity in maize hybrids is crucial for fumonisin contamination in kernels. *Journal of Cereal Science* 54: 467-472.
- Battilani, P., Pietri, A., Barbano, C., Scandolara, A., Bertuzzi, T. and Marocco, A., 2008. Logistic regression modeling of cropping systems to predict fumonisin contamination in maize. *Journal of Agricultural and Food Chemistry* 56: 10433-10438.
- Chungu, C., Mather, D.E., Reid, L.M. and Hamilton, R.I., 1996. Comparison of techniques for inoculating maize silk, kernel, and cob tissues with *Fusarium graminearum*. *Plant Disease* 80: 81-84.
- De Mendiburu, F., 2017. *Agricolae: statistical procedures for agricultural research*. Available at: <https://CRAN.R-project.org/package=agricolae>
- Desjardins, A.E. and Proctor, R.H., 2007. Molecular biology of *Fusarium* mycotoxins. *International Journal of Food Microbiology* 119: 47-50.
- Dorn, B., Forrer, H.R., Jenny, E., Wettstein, F.E., Bucheli, T.D. and Vogelgsang, S., 2011. *Fusarium* species complex and mycotoxins in grain maize from maize hybrid trials and from grower's fields. *Journal of Applied Microbiology* 111: 693-706.
- Eller, M., Robertson-Hoyt, L.A., Payne, G.A. and Holland, J.B., 2008. Grain yield and *Fusarium* ear rot of maize hybrids developed from lines with varying levels of resistance. *Maydica* 53: 231-237.
- Filipovic, M., Babic, M., Delic, N., Bekavac, G. and Babic, V., 2014. Determination of relevant breeding criteria by the path and factor analysis in maize. *Genetika* 46: 49-58.
- Food and Agriculture Organization of the United Nations (FAO), 2017. *The future of food and agriculture – trends and challenges*. FAO, Rome, Italy.
- Food and Agriculture Organization of the United Nations (FAO), 2018. *FAOSTAT statistics database collections*. FAO, Rome, Italy.
- Gaikpa, D.S. and Miedaner, T., 2019. Genomics-assisted breeding for ear rot resistances and reduced mycotoxin contamination in maize: methods, advances and prospects. *Theoretical and Applied Genetics* 132: 2721-2739.
- Guo, B., Ji, X., Ni, X., Fountain, J.C., Li, H., Abbas, H.K., Lee, R.D. and Scully, B.T., 2017. Evaluation of maize inbred lines for resistance to pre-harvest aflatoxin and fumonisin contamination in the field. *Crop Journal* 5: 259-264.
- Henry, W.B., Williams, W.P., Windham, G.L. and Hawkins, L.K., 2009. Evaluation of maize inbred lines for resistance to *Aspergillus* and *Fusarium* ear rot and mycotoxin accumulation. *Agronomy Journal* 101: 1219-1226.
- Hope, R.M., 2013. *Rmisc: Rmisc: Ryan Miscellaneous*. R package version 1.5. Available at: <https://CRAN.R-project.org/package=Rmisc>
- Hung, H. and Holland, B.J., 2012. Diallel analysis of resistance to *Fusarium* ear rot and fumonisin contamination in maize. *Crop Science* 52: 2173-2181.
- Ju, M., Zhou, Z., Mu, C., Zhang, X., Gao, J., Liang, Y., Chen, J., Wu, Y., Li, X., Wang, S., Wen, J., Yang, L. and Wu, J., 2017. Dissecting the genetic architecture of *Fusarium verticillioides* seed rot resistance in maize by combining QTL mapping and genome-wide association analysis. *Scientific Reports* 7: 46446.
- King, S.B. and Scott, G.E., 1982. Field inoculation techniques to evaluate maize for reaction to kernel infection by *Aspergillus flavus*. *Phytopatology* 72: 782-785.
- Lanubile, A., Ferrarini, A., Maschietto, V., Delledonne, M., Marocco, A. and Bellin, D., 2014b. Functional genomic analysis of constitutive and inducible defense responses to *Fusarium verticillioides* infection in maize genotypes with contrasting ear rot resistance. *BMC Genomics* 15: 710.
- Lanubile, A., Logrieco, A., Battilani, P., Proctor, R.H. and Marocco, A., 2013. Transcriptional changes in developing maize kernels in response to fumonisin-producing and nonproducing strains of *Fusarium verticillioides*. *Plant Science* 210: 183-192.
- Lanubile, A., Maschietto, V. and Marocco, A., 2014a. Breeding maize for resistance to mycotoxins. In: Leslie, J.F. and Logrieco, A.F. (eds.) *Mycotoxin reduction in grain chains*. John Wiley & Sons, Ltd., Chichester, UK, pp. 37-58.

- Lanubile, A., Maschietto, V., Battilani, P. and Marocco, A., 2017b. Infection with toxigenic and atoxigenic strains of *Aspergillus flavus* induces different transcriptional signatures in maize kernels. *Journal of Plant Interactions* 12: 21-30.
- Lanubile, A., Maschietto, V., Borrelli, V.M., Stagnati, L., Logrieco, A.F. and Marocco, A., 2017a. Molecular basis of resistance to *Fusarium* ear rot in maize. *Frontiers in Plant Science* 8: 1774.
- Lanubile, A., Maschietto, V., De Leonardi, S., Battilani, P., Paciolla, C. and Marocco, A., 2015. Defense responses to mycotoxin producing fungi *Fusarium proliferatum*, *F. subglutinans*, and *Aspergillus flavus* in kernels of susceptible and resistant maize genotypes. *Molecular Plant-Microbe Interaction* 28: 546-557.
- Lanubile, A., Pasini, L. and Marocco, A., 2010. Differential gene expression in kernels and silks of maize lines with contrasting levels of ear rot resistance after *Fusarium verticillioides* infection. *Journal of Plant Physiology* 167: 1398-406.
- Lanubile, A., Pasini, L., Lo Pinto, M., Battilani, P., Prandini, A. and Marocco, A., 2011. Evaluation of broad spectrum sources of resistance to *Fusarium verticillioides* and advanced maize breeding lines. *World Mycotoxin Journal* 1: 43-51.
- Marin, S., Ramos, A.J., Cano-Sancho, G. and Sanchis V., 2013. Mycotoxins: occurrence, toxicology, and exposure assessment. *Food and Chemical Toxicology* 60: 218-237.
- Maschietto, V., Colombi, C., Pirona, R., Pea, G., Strozzi, F., Marocco, A., Rossini, L. and Lanubile, A., 2017. QTL mapping and candidate genes for resistance to *Fusarium* ear rot and fumonisin contamination in maize. *BMC Plant Biology* 17: 20.
- Maschietto, V., Lanubile, A., Leonardi, S.D., Marocco, A. and Paciolla, C., 2016. Constitutive expression of pathogenesis-related proteins and antioxidant enzyme activities triggers maize resistance towards *Fusarium verticillioides*. *Journal of Plant Physiology* 200: 53-61.
- Mauro, A., Battilani, P., Callicott, K.A., Giorni, P., Pietri, A. and Cotty, P.J., 2013. Structure of an *Aspergillus flavus* population from maize kernels in northern Italy. *International Journal of Food Microbiology* 162: 1-7.
- Mesterházy, Á., Lemmens, M. and Reid, L.M., 2012. Breeding for resistance to ear rots caused by *Fusarium* spp. in maize – a review. *Plant Breeding* 131: 1-19.
- Morales, L., Marino, T.P., Wennadt, A.J., Fouts, J.Q., Holland, J.B. and Nelson, R.J., 2018. Dissecting symptomatology and fumonisin contamination produced by *Fusarium verticillioides* in maize ears. *Phytopathology* 108: 1475-1485.
- Munkvold, G., 2003. Epidemiology of *Fusarium* diseases and their mycotoxins in maize ears. *European Journal of Plant Pathology* 109: 705-713.
- Nielsen, R.L., Thomison, P.-R., Brown, G.A., Halter, A.L., Wells, J. and Wuethrich, K.L., 2002. Delayed planting effects on flowering and grain maturation of dent corn. *Agronomy Journal* 94: 549.
- O'Brian, G.R., Georgianna, D.R., Wilkinson, J.R., Yu, J., Abbas, H.K., Bhatnagar, D., Cleveland, T.E., Nierman, W. and Payne, G.A., 2007. The effect of elevated temperature on gene transcription and aflatoxin biosynthesis. *Mycologia* 99: 232-239.
- Okoth, S., Rose, L.J., Ouko, A., Netshifhephe, N.E.I., Sila, H. and Viljoen, A., 2017. Assessing genotype by environment interactions in *Aspergillus* ear rot and preharvest aflatoxin accumulation in maize inbred lines. *Agronomy* 7: 86.
- R Core Team, 2017. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Reid, L.M., Zhu, X., Parker, A. and Yan, W., 2009. Increased resistance to *Ustilago zea* and *Fusarium verticillioides* in maize inbred lines bred for *Fusarium graminearum* resistance. *Euphytica* 165: 567-578.
- Robertson, L.A., Kleinschmidt, C.E., White, D.G., Payne, G.A., Maragos, C.M. and Holland, J.B., 2006. Heritabilities and correlations of *Fusarium* ear rot resistance and fumonisin contamination resistance in two maize populations. *Crop Science* 46: 353-361.
- Robertson-Hoyt, L.A., Betrán, J., Payne, G.A., White, D.G., Isakeit, T., Maragos, C.M., Molnár, T.L. and Holland, J.B., 2007. Relationships among resistances to *Fusarium* and *Aspergillus* ear rots and contamination by fumonisin and aflatoxin in maize. *Phytopathology* 97: 311-317.
- Rose, L.J., Okoth, S., Beukes, I., Ouko, A., Mouton, M., Flett, B.C., Makumbi, D. and Viljoen, A., 2017. Determining resistance to *Fusarium verticillioides* and fumonisin accumulation in maize inbred lines resistant to *Aspergillus flavus* and aflatoxins. *Euphytica* 213: 93.
- Santiago, R., Cao, A. and Butrón, A., 2015. Genetic factors involved in fumonisin accumulation in maize kernels and their implications in maize agronomic management and breeding. *Toxins* 7: 3267-3296.
- Scott, G.E. and Zummo, N., 1990. Registration of Mp313E parental line of maize. *Crop Science* 30: 1378.
- Septiani, P., Lanubile, A., Stagnati, L., Busconi, M., Nelissen, H., Pè, M.E., Dell'Acqua, M. and Marocco, A., 2019. Unravelling the genetic basis of *Fusarium* seedling rot resistance in the MAGIC maize population: novel targets for breeding. *Scientific Reports* 9: 5665.
- Stagnati, L., Lanubile, A., Samayoa, L.F., Bragalanti, M., Giorni, P., Busconi, M., Holland, J.B. and Marocco, A., 2019. A genome wide association study reveals markers and genes associated with resistance to *Fusarium verticillioides* infection of seedlings in a maize diversity panel. *G3: Genes, Genomes, Genetics* 9: 571-579.
- Szabo, B., Toth, B., Toth Toldine, E., Varga, M., Kovacs, N., Varga, J., Kocsube, S., Palagyi, A., Bagi, F., Budakov, D., Stojšin, V., Lazić, S., Bodroža-Solarov, M., Čolović, R., Bekavac, G., Purar, B., Jocković, D. and Mesterházy, A., 2018. A new concept to secure food safety standards against *Fusarium* species and *Aspergillus flavus* and their toxins in maize. *Toxins* 10: 372.
- Venables, W.N. and Ripley, B.D., 2002. Modern applied statistics with S, 4th edition. Springer, New York, NY, USA.
- Warburton, M.L., Williams, W.P., Windham, G.L., Murray, S.C., Xu, W., Hawkins, L.K. and Duran, J.F., 2013. Phenotypic and genetic characterization of a maize association mapping panel developed for the identification of new sources of resistance to *Aspergillus flavus* and aflatoxin accumulation. *Crop Science* 53: 2374-2383.
- Zila, C.T., Ogut, F., Romay, M.C., Gardner, C.A., Buckler, E.S. and Holland, J.B., 2014. Genome-wide association study of *Fusarium* ear rot disease in the U.S.A. maize inbred line collection. *BMC Plant Biology* 14: 372.
- Zila, C.T., Samayoa, L.F., Santiago, R., Butrón, A. and Holland, J.B., 2013. A genome-wide association study reveals genes associated with *Fusarium* ear rot resistance in a maize core diversity panel. *G3: Genes, Genomes, Genetics* 3: 2095-2104.
- Zuber, M.S., 1973. Registration of 20 maize parental lines. *Crop Science* 13: 779.

